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Penicillin-Induced Agranulocytosis

BARBARA WALZ, MD JOSEPH ZUPNICK, MD DAVID UDEN, MD ARMAND KEATING, MD Toronto

Systemic penicillin therapy is generally the treatment of choice in patients with bacterial endocarditis. Prolonged therapy at a high dosage is often required. Complications of treatment with penicillin, although uncommon, include hypersensitivity reactions ranging from mild urticaria to lifethreatening anaphylaxis, central nervous system disorders, electrolyte disturbances, and acute interstitial nephritis. Hematologic complications of hypersensitivity to penicillin are rare. Coombs' test-positive hemolytic anemia has been reported.1,2 Leukopenia also occurs, with 26 cases being reported to date. Agranulocytosis, however, has been previously reported only once, in 1946.3 Herein we report a second case of agranulocytosis as a complication of penicillin therapy.

Report of a Case

The patient, a 17-year-old woman, presented with a three-week history of nocturnal fever not associated with rigors or chills. She had had generalized myalgia for two weeks and a generalized maculopapular rash for one week. Her family physician recorded a mitral regurgitant murmur and referred her to our hospital. There was no history of rheumatic fever, and her last dental visit was three months before the symptoms began.

On physical examination on admission she had a grade III/VI pansystolic murmur at the apex and a maculopapular rash over her extremities, but the results were otherwise normal. A chest x-ray film and an electrocardiogram were normal. Two-dimensional echocardiograms showed mitral valve prolapse with no valvular vegetations.

A clinical diagnosis of infective endocarditis was made and confirmed by identifying viridans group streptococci in each of nine blood cultures. The organism was sensitive to penicillin in vitro. Laboratory findings included the following: hemoglobin, 134 grams per liter; leukocyte count, 16.9×10^9 per liter, with 62% neutrophils and 24% lympho-

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cytes. The urea, creatinine, electrolytes, plasma proteins, and liver enzyme levels were normal.

The intravenous administration of penicillin G sodium, 3 million units every six hours, and streptomycin sulfate, 100 mg given intramuscularly daily, was begun. Five days after starting treatment the patient was asymptomatic. On day 17 of her hospital stay, the streptomycin therapy was discontinued. The patient continued to do well until day 23, when she complained of a sore throat. Her leukocyte count was 3.2 $\times 10^9$ per liter. Neutrophils were absent; the nucleated cells were predominantly atypical lymphocytes. Her hemoglobin was 130 grams per liter and the platelet count 332×10^9 per liter. A Coombs' test and assays for hepatitis B-associated antigen, heterophile antibody, antinuclear antibody, and rheumatoid factor were all negative. Serum immunoelectrophoresis showed an increase in polyclonal immunoglobulin (Ig) M and IgG. C3, C4, and CH₅₀ assays were all negative.

A bone marrow aspirate showed normal cellularity with normal erythropoiesis. The myeloid:erythroid ratio was 3:1. Granulopoiesis showed no maturation beyond the promyelocyte stage.

Treatment with penicillin was discontinued on day 25, and a regimen of vancomycin hydrochloride was started. Two days later a peripheral blood monocytosis arose, accompanied by the appearance of bands. By day 30 the leukocyte count had risen to 17×10^9 per liter, with 0.48 neutrophils, 0.15 bands, 0.29 lymphocytes, and 0.08 monocytes. On discharge from hospital on day 34, there was no clinical evidence of the original infection.

Discussion

Leukopenia due to penicillin therapy is a rare event; only 26 cases—including this one—have been reported to date. We believe this to be the second report of penicillin-induced agranulocytosis in the past 50 years. In each case of penicillin-associated leukopenia, the patients received high-dosage penicillin for serious systemic infections. In 18 of 26 cases, therapy was for endocarditis. Table 1 summarizes the cases reported to date. All patients received their drugs by the intravenous or intramuscular routes. The earliest that leukopenia was noted was after four days of therapy, although it was seen as late as 35 days after penicillin treatment in one patient. The average duration of therapy before leukopenia occurred was 20 days. The range of total leukocyte counts at the nadir of the leukopenia was 0.1 to 3.9×10^9 per liter. Including our case, the absolute granulocyte count ranged from 0 to 1.998×10^9 per liter. The average granulocyte count was 0.372×10^9 per liter. In keeping with other reports, the total dose at the start of leukopenia was greater than 150,000 units per kg body weight per day. All patients had normal leukocyte counts within 2 to 14 days of discontinuing the antibiotic therapy. The average time to recovery was 4.7 days. There were no deaths or permanent sequelae related to the leukopenia. Five patients had Coombs' test-positive hemolytic anemia. 1,2,5-7

Bone marrow aspiration was done in 16 cases. The results of the aspirations showed no consistent pattern. Hypercellular, normocellular, and hypocellular marrows have been reported, although a hypocellular marrow was found in only one case.11 In this case of gonococcal endocarditis, there appeared to be no features distinguishing it from cases associated with hypercellular and normocellular marrows.

Our case is consistent with other reports in total penicillin

From the Divisions of Hematology and Cardiology, Toronto Western Hospital,

Reprint requests to Armand Keating, MD, Oncology Clinic, Toronto General Hospital, 200 Elizabeth St, Toronto, Ontario M5G 2C4

dose, duration of treatment, and duration of leukopenia. While it is possible that the leukopenia may have resulted from sepsis alone, it is more likely that the common factor was high doses of parenteral penicillin. The mechanisms of penicillin-induced leukopenia are as yet not fully understood. At least two distinct mechanisms of drug-induced leukopenia are known,^{6.17} and both immunologic (type I) and toxic (type II) factors have been implicated. Antineutrophil

*Leukopenia recurred with the readministration of penicillin.

†Patient had a previous episode of penicillin G-induced granulocytopenia

antibodies have been detected in seven patients, 5.6.15.16 and circulating antipenicillin antibodies have been described as well. 2.6.18 Moreover, in a case described by Rossiter and co-workers, there was a previous history of penicillin-induced leukopenia. 5 Similarly, in the case described by Petz and Fudenberg, the readministration of penicillin resulted in a recurrence of Coombs' test-positive hemolytic anemia with leukopenia. 2 Interestingly, the initial hemolytic anemia with

	Source	Diagnosis		Sex	Penicillin Dose/d, million units	Days to	Cell Co	ounts at	Nadir				
_			<u> </u>			Onset of Leuko- penia	WBC PMN %		Days to	Anti- leukocyte			
Case							×	10 ⁹ /lite	er 	Recovery	Antibody	Test	Bone Marrow
	Spain and Clark, 1946 ³	Bowel obstruction	54	O'	100,000 U × 1d 160,000 U × 3d	4	0.10	0.00	0	1			Normal at autopsy
	Petz and Fudenberg, 1966 ²	Infective endocarditis	45	O*	100 IV	23	1.60	0.18	11	7		Pos	Normal cellularity; decreased granulocyte precursors
3	Forshaw, 19688	Infective endocarditis	47	O*	8 IV × 14d 16 IV × 6d 32 IV × 13d	27	0.95	0.02	2	14	• • •	Neg	Increased cellularity; no mature granulocytes
	Rossiter et al, 1968 ⁵	Infective endocarditis	37	ď	Phenethicillin, 4 grams PO × 14d, then IM	28	0.80	0.03	4	3	Pos	Pos	Normal erythropoiesis; no mature granulocyte
5	White et al, 1968 ¹	Infective endocarditis	64	Q	20 IV	18	3.20			2		Pos	Increased erythropoies left-shifted granulopoie
	Joorabchi and Kohout, 1973 ⁷	Infective endocarditis	12	O*	20 IV	18	2.30	0.16	7	14		Pos	Hypercellular, M:E 3:1, shift toward mature ce
	Colvin et al, 1974 ⁹		39	Q	20 IV × 7d 12 IV × 14d	23	0.90	0.08	9	5	Neg	Neg	Hypercellular; increase granulopoiesis
	Homayouni et al, 19794	Cellulitis	60	o*	230,000 U/kg IV	13	3.70	1.99	54	6	• • •		
9		Gangrenous appendix	81	O*	250,000 U/kg IV	9	3.70	1.81	49	2	• • •	• • •	•••
10‡	Neftel et al, 1981 ⁶	Infective endocarditis	54	Q	12 IV × 18d 32 IV × 3d	20	0.80	0.04	2	2			
11‡		Infective endocarditis	28	O*	24 IV	19	2.00	0.10	5	4	Pos	Pos	• • •
12‡		Infective endocarditis	27	O*	16 IV	21	2.20	0.23	10	2			Hypocellular; maturation arrest, promyelocyte stage
13‡		Infective endocarditis	39	O*	20 IV	26	2.20		•••	6			
14‡		Infective endocarditis	25	ď	20 IV	24	1.80	• • •	• • •	3	Pos		Hyperactive marrow
15‡		Pneumonia	35	O*	16 IV	26	3.90	• • •	• • •	4	Pos		Maturation arrest, myelocyte stage
16		Hemangioma	13	O*	12 IV	19	3.40			10	Pes	• • •	Maturation arrest, myelocyte stage
17‡		Septic arthritis	64	Q	20 IV	26	2.50	1.00	40	3			• • •
	Postelnick and Gaskins, 1981 ¹⁰	Infective endocarditis	21	Q	16 IV × 14d 20 IV × 3d	15	1.80	0.09	5	2	• • •		• • •
19§	Timmis et al, 1981 ¹¹	Infective endocarditis	18	Q	20 IV	14	2.60	0.21	8	7			Hypoplastic
20	Yap, 1981 ¹²	Infective endocarditis	32	O'	16 IV × 7d 8 IV × 18d	25	1.20	0.21	17	3	• • •	Neg	
21	Corbett et al, 1982 ¹³	Infective endocarditis	28		8 × 20d 13 × 1d	22		0.40		3		• • •	Hypercellular; left-shif granulopoiesis
22		Infective endocarditis	55		12	35		0.50		2	• • •		Maturation arrest of WBC series
	Miro et al, 1983 ¹⁴	Pleural empyema	40	o•	12	24		0.31		4		• • •	
24	Rouveix et al, 1983 ¹⁵	Infective endocarditis	46		250,000 U/kg IV	25		0.02	1	4	Pos		
25	Snavely et al, 1983 ¹⁶	Infective endocarditis	31	ď	6 IV × 15d 15 IV × 14d	29	2.20	0.06	3	5	Pos	Neg	Hypercellular; increas granulocyte series; paucity of mature for
26	This report, 1988	Infective	17	^	12 IV	22	2 20	0.00	0	8	Neg	Neg	Maturation arrest,

±Data extrapolated

§Granulocytopenia recurred after cefuroxime sodium and gentamicin sulfate were given.

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leukopenia developed in this patient despite the prophylactic administration of prednisone. In another case where no antibodies were detected, the readministration of a single dose of penicillin resulted in a striking drop in the neutrophil count within 24 hours.7 Although Neftel and associates have described circulating antipenicillin IgG antibodies in all their patients given penicillin in cumulative doses greater than 200 million units, 6,18 leukopenia did not develop in all patients so sensitized. The degree of sensitization may be diminished if the drug is given as a freshly prepared bolus rather than by slow infusion. 18,19

The reports of neutropenia with the readministration of penicillin and the presence of antipenicillin antibodies favor a type I or hypersensitivity-type reaction. If this postulate is correct, this leaves the question of why antineutrophil antibodies have not been detected more frequently. One possibility is that the antibodies are of low concentration and of high affinity. In this situation, the antibodies would be almost all cell-bound, hence the free, detectable amount in the circulation would be low. If penicillin degradation products are required as haptens for the neutrophils, these would not be detected if fresh penicillin were added to the in vitro assay.14 The occurrence, however, of penicillin-associated neutropenia in the face of prednisone therapy¹ argues against an immune-mediated reaction.

Another possible mechanism is that of a toxic, doserelated suppression of bone marrow precursors. This mechanism would be independent of circulating antipenicillin or antineutrophil antibodies. The almost invariable association with the administration of high doses of the drug and the fact that the neutropenia seldom occurred before 14 days favor this concept. Alternatively, the high dose and delayed onset may be related to the minimum dose needed for sensitization to occur. Joorabchi and Kohout have postulated that the pancytopenia seen in their patient was due to the penicillininduced blockade of the release of mature cells from the marrow.7

The variation of marrow findings in the reported cases is intriguing and could possibly be explained by variations in the time of marrow sampling. Early in a patient's course, the marrow may be hypoplastic or aplastic and devoid of granulocytes and normoblasts. During the earliest phase of marrow recovery, there may be a transient increase in small dense lymphocytes in the marrow, subsequently replaced by the earliest granulocyte precursors. ¹⁷ Later, during recovery, the marrow would be extremely hypercellular with intense mitotic activity. Thus, "maturation arrest"—in which differentiation of marrow cells is considered to halt at a specific developmental stage—may be related only to the phase of recovery of the regenerating marrow.

Leukopenia is a rare complication of systemic penicillin therapy. It almost invariably occurs in cases of prolonged high-dose therapy. Monitoring the leukocyte count regularly during therapy may avert the occurrence of absolute granulocytopenia by detecting leukopenia early. Withdrawing penicillin therapy has resulted in a rapid return to normal of the leukocyte count in all cases.

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Midcycle Pneumothorax in a Patient With Catamenial **Pneumothoraces**

ANDREW J. GHIO, MD ROBERT O. CRAPO, MD Salt Lake City

CATAMENIAL PNEUMOTHORAX is a syndrome of recurrent pneumothoraces developing within 48 to 72 hours of the start of menstrual bleeding. In most cases they are small, unilateral, right-sided pneumothoraces occurring in parous women in their third and fourth decades. The close temporal relationship to menses has led several authors to conclude that evidence of pneumothorax at other times in the cycle precludes the diagnosis. 1-3 We report the case of a patient with catamenial pneumothorax in whom pneumothoraces developed between menses.

Report of a Case

The patient, a 39-year-old nonsmoking woman, presented for evaluation of recurrent pneumothoraces. She had been admitted to hospital twice with shortness of breath, cough, and chest pain. Chest radiographs showed small right-sided pneumothoraces without other abnormalities. During both hospital stays, therapy included only oxygen and analgesics. She recalled that each admission corresponded with the beginning of her menstrual period. In the following year the same symptoms recurred several times, always within 48 hours of the start of menstrual bleeding, but she saw her physician only when she needed analgesics. On each visit, a chest radiograph showed a small right-sided pneumo-

A year after her initial diagnosis, she also noticed shortness of breath, cough, and chest pain between menses. On day 14 of a menstrual cycle, she consulted her physician at the

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Reprint requests to Andrew J. Ghio, MD, Pulmonary Division, LDS Hospital, Salt Lake City, UT 84143.

From the Pulmonary Division, Department of Medicine, LDS Hospital and the University of Utah School of Medicine, Salt Lake City.